Sensitive Molecular Identification of Pathogens causing Implant and Tissue Infections (ITI)

K. Hohenwarter¹, E. Scherfler¹, W. Prammer¹, W. Aichinger¹, S. Eisenberger², B. Ronacher²

Klinikum Wels-Grieskirchen, Wels, AUSTRIA

² Cube Dx GmbH, St. Valentin, AUSTRIA

Objectives

Prosthetic infection is the most severe complication in joint arthroplasty. The diagnostic procedure is time consuming and in many cases unrewarding. Microbial growth can be slowed or suspended if the pathogens are weakened by antimicrobial therapy. Molecular diagnostics is a reliable complement for optimizing conventional microbiology by detecting bacteria that do not grow in culture. We used compact sequencing, a combination of highly-sensitive Polymerase Chain Reaction (PCR) with hybcell based identification, to detect and identify pathogenic bacteria or fungi in clinical tissue and synovial fluid samples. These samples were tested in parallel to bacterial culture in combination with identification by MALDI-TOF. Our aim was to test the suitability of compact sequencing to detect and identify pathogens as an additional standard method to diagnose implant or tissue infections.

Method

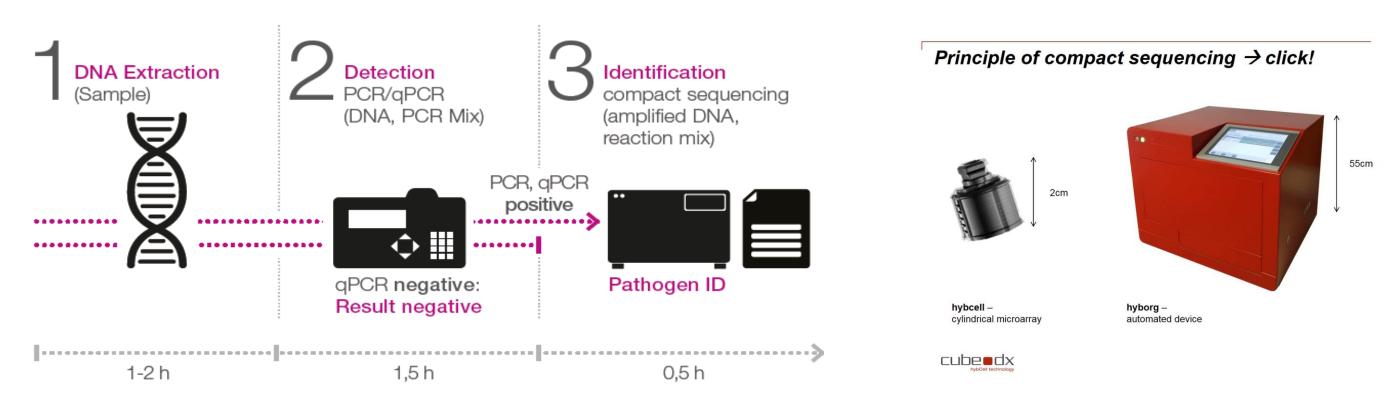
52 samples were tested: 40 samples from joints (shoulder, elbow, hip and knee), 4 from ascites and 8 from other sources. Samples from infected tissue were divided into two aliquots. One was used for bacterial culturing on agar and in blood culture bottles for 6 weeks. The second aliquot was stored deep frozen (- 80° C) and later used for pathogen DNA extraction with SelectNA (Molzym, DE). The detection and identification of bacteria and fungi were done by compact sequencing based on hybcell ITI DNA xA (Cube Dx, AT).

Results

The Cube Dx test shows a sensitivity of 93% and a specificity of 63%, a negative and positive predictive value of 96% and 50%, respectively and an accuracy of 71% in comparison to microbiological evaluation. The total number of samples testing positive for bacteria and fungi was higher for compact sequencing with 53% (28 of 52 samples) than for bacteriological culture with 29% (15 of 52 samples).

Conclusion

Our results demonstrate a good correlation between molecular and cultural detection. In 27% (14 of 52 samples) of all cases, molecular testing was more sensitive than culture. hybcell ITI DNA xA is well suited for use as an additional sensitive diagnostic tool for critical clinical samples in a routine lab.



Workflow of hybcell ITI DNA xA.



ECCMID

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| No. | Source | Culture | hybcell ITI DNA xA | |
|----------|-------------------------|-------------------------------|------------------------------------------------------------|--|
| | | | Streptococcus anginosus; | |
| 1 | abdominal cavity | Candida albicans | Klebsiella oxytoca, Candida albicans | |
| | | Pseudomonas aeruginosa, | | |
| 2 | abscess cavity | Enterococcus faecalis, | Bacteria pan | |
| - | | Bacteroides fragilis, | | |
| | have a la se a da | Bacteroides thetaiothaomicron | | |
| 3 | lymph node | Streptococcus salivarius | Streptococcus (genus) | |
| 4 | secretion from | Coagulase negative | Staphylococcus haemolyticus, Anaerococcus (genus), | |
| 4 | urethra | staphylococcae | Corynebacterium (genus) | |
| 5 | serom latissimus | neg | Nectriaceae | |
| 6 | upper arm | Staphylococcus aureus | Staphylococcus aureus | |
| 7 | upper arm | Staphylococcus aureus | Staphylococcus aureus | |
| 8 | ascites | neg | Bacteria pan | |
| 9 | ascites | neg | neg | |
| 10 | ascites | neg | neg | |
| 11 | elbow | Staphylococcus aureus | Staphylococcus aureus | |
| 12 | elbow, bursa | neg | neg | |
| 13 | hip joint | neg | neg | |
| 14 | knee joint | Candida albicans | Enterococcus faecalis, Candida albicans | |
| 15 | knee joint | Staphylococcus aureus | Staphylococcus aureus | |
| 16 | knee joint | Staphylococcus aureus | Staphylococcus aureus | |
| 17 | knee joint | Streptococcus pyogenes | Streptococcus pyogenes | |
| 18 | knee joint | Streptococcus pyogenes | Streptococcus pyogenes | |
| 19 | knee joint | Corynebacterium ssp. | neg Anaerococcus (genus), | |
| 20 | knee joint | neg | Corynebacterium (genus) | |
| 21 | knee joint | neg | neg | |
| 22 | knee joint | neg | Candida (genus) | |
| 23 | knee joint | neg | Streptococcus (genus) | |
| | | | Anaerococcus (genus), | |
| 24 | knee joint | neg | Corynebacterium (genus) | |
| 25 | shoulder joint | Staphylococcus epidermidis | Staphylococcus (genus) | |
| 26 | shoulder joint | neg | neg | |
| 27 | knee joint | neg | neg | |
| 28 | knee joint | neg | Staphylococcus aureus | |
| 29 | knee joint | neg | Staphylococcus aureus | |
| 30 | knee joint | neg | neg | |
| 31 | knee joint | neg | neg | |
| 32 | knee joint | neg | neg | |
| 33 34 | knee joint | neg | neg | |
| | perikard | neg | neg Candida (genus), Candida dubliensis, | |
| 35 | knee joint | Candida parapsilosis | Candida (genus), Candida dubilensis, Candida tropicalis | |
| 36 | knee joint | neg | Streptococcus (genus) | |
| 37 | hip joint | neg | neg | |
| 38 | knee joint | neg | neg | |
| 39 | shoulder joint | neg | neg | |
| 40 | knee joint | neg | Bacteria pan | |
| 41 | knee joint | neg | neg | |
| 42 | knee joint | neg | Streptococcus (genus) | |
| 43 | knee joint | neg | neg | |
| 44 | knee joint | neg | Candida (genus) | |
| 45 | knee joint | neg | neg | |
| 46 | knee joint | neg | neg | |
| 47 | shoulder joint | neg | neg | |
| 48 | knee joint | neg | neg | |
| 49 | aszites | neg | Bacteria pan | |
| 50 51 | hip joint knee joint | neg | Streptococcus agalactiae | |
| 51 | shoulder joint | neg | neg neg | |
| JZ | shoulder joint | neg | neg | |

| | | ture | | |
|---------|------|------|------|----|
| | | pos. | neg. | Σ |
| cell | pos. | 14 | 14 | 28 |
| hybcell | neg. | 1 | 23 | 24 |
| | Σ | 15 | 37 | 52 |

Results and correlation of culture and hybcell.